

# Health Benefits of Pomegranate (Peel & Juice) and Preparation of Functional Pomegranate Drink Using Probiotic *Lactobacillus Plantarum*

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## Original Article

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## ABSTRACT

Pomegranate (*Punica granatum L.*) is rich in bioactive compounds, antioxidant, and volatile substances; under this investigates the impact of pomegranate peel (wastes), juice and preparation of probiotic juice by *Lactobacillus plantarum* on antimicrobial activity, antidiabetic *in vitro* and hypocholesterolemic agents by rats. Pomegranate peel extract has the highest total phenol, flavonoid and antioxidant more than juice .but no significance with probiotic juice. Antimicrobial compounds from pomegranate peel and probiotic juice are effective against a range of pathogenic microorganisms. The anti-amylase activity was significantly influenced by pomegranate tissues. The biological experiment used (n=36 rats up to 6 weeks) which were divided into six groups G1 (- ve control). G2 (positive control) by high-fat diet + 1% cholesterol, G3 (positive control+ 300 mg/kg/day/rat pomegranate peel extract), G4 (positive control+ 1 ml juice / day/rat), G5 (positive control+1 ml probiotic juice / day/rat), and G6 (positive control + reference drug 0.18 mg/kg/day/rat (ATOR 20 mg). Results, on the other hand, demonstrate the impact of peel extract, juice, and probiotic juice on lipid content (LDL, HDL, total cholesterol and triglycerides) and liver-related enzymes (ALT, ALP, AST). The pomegranate peel extracts and probiotic juice is more effective than juice due to the rich phenolic content and fermentation of juice by the probiotic strain *Lactobacillus plantarum*.

## 1. Introduction

The pomegranate, or *Punica granatum L.*, is a very old fruit that has been consumed extensively throughout history by many different cultures. It is regarded as a super fruit with numerous health-improving qualities, including antimicrobial, antiallergic, anti-tumor, and antioxidant (Alkhatib et al., 2022). Pomegranate which contains several phytochemicals, each of which has more than one bioactivity, has various medicinal uses, such as being antibacterial, antiviral, and antifungal. It has been used since ancient times until now, according to (Stefanou et al., 2020). Pomegranate is a fruit with a biowaste (peel) that has the ability to be transformed into added value for a wide range of products. Which use pomegranate rind has antioxidant and antibacterial effects which able to inhibit the main path-

ogenic microorganism (Ko et al., 2021 and Elzoghbiy et al., 2022) and Xiang (Luo et al., (2020) reported that *Punica granatum L.* cultivation dates back to the dawn of civilization, and since the turn of the 20<sup>th</sup> century, as the fruit's health advantages have been confirmed by science, production and consumption have expanded. Pomegranate fruit peels are one of the main byproducts that are highly concentrated in broad-spectrum antioxidants and antibacterial agents; they can even stop food from going bad. Pomegranate is useful for food preservation and possesses antibacterial properties, making it a complete resource for farmers, the food processing and storage industries. Probiotic bacteria have been used as food additives that improve the balance of intestinal microflora, however dairy products are now the best frequent food carriers to reliable

source of probiotics, an raising variety of non-dairy food substrates display the ability for transmission of probiotics, according to studies on the functional food industry (Min et al., 2019).

Probiotic bacteria appear to be used in the fermentation of juice to produce beverages with high nutritional benefits and regarded organoleptic quality (Stavros et al., 2020). Due to their improved nutritional value, sensory qualities, improved storage life, and addition of derived components that promote health, fermented fruit beverages serve functional manufacturing needs (Hua et al., 2021).

(Amri et al., 2020) shows that leaves, rind, or juice extracts of pomegranate promote decrease insulin sensitivity and promote glycemic control, and limit carbohydrate and lipid absorption by lowering the activities of amylase and lipase. (Andi Alfira et al., 2020) studied In Vitro that the peel powder of fruit Pomegranate had great effect on amylase inhibitor. (Seba et al., 2021) assayed that the pomegranate, seeds ,peel, meso-carp and juice displayed higher inhibiting behavior on  $\alpha$ -glucosidase and  $\alpha$ -amylase, this is definitely connected to a lower IC<sub>50</sub> value. The enzyme that causes the postprandial glucose level in diabetics to increase is due to the release of sugar molecules (oligosaccharides and monomers) from eating complex carbohydrates is alpha-amylase, which plays a role in regulating blood plasma glucose levels. The plan was implemented to study the chemical, microbiological, and biological activity of pomegranate (peel, juice, and probiotic juice).

## 2. Materials and Methods

### Materials

#### Plant materials

One variety of pomegranates (*Punica granatum L.*) Manfaloty was received from the Agricultural Research Center, Giza, Egypt's Horticulture Research Institute, at their full maturity. The pomegranates were rinsed thoroughly with distilled water after being thoroughly cleaned under running tap water, and then left to dry at room temperature in the air for a few minutes.

#### Pomegranate peel powder preparation

Peels were chopped into bits. At a temperature

of 45°C for 72 hours, the peel was dried in the tray dryer. The grinder was used to process dried pomegranate peels into smaller particles. Thereafter, a sieve with a mesh size of 50 is used to catch the ground particles. Then stored at ambient temperature (4±1°C) until use (Devatkal and Naveena, 2010).

#### Pomegranate juice preparation

The pomegranate fruits and peeled off to create pure juice, the seeds were manually separated and crushed in a blender. The extraction step was put through the juicer once again to extract more juice then filtered through muslin cloth and kept at 4°C until use. 25 ml of a 50% (v/v) solution of water and ethanol were added to 5 g of juice. The peel powder was processed using the same method; the extracts were filtered and concentrated using a powerful evaporator (Eyela rotary vacuum evaporator N-11; Tokyo ridadidai Co., Ltd., Japan) (Gözlekçi et al., 2011).

#### Preparation of starter culture

*Lactobacillus plantarum* was starter culture which was cultivated 24h at 37°C in the MRS broth. 10 mL of the individually grown MRS broths at 4000 rpm for 10 min were centrifuged to get the biomass. To remove the remaining MRS culture, the resulting biomass was twice cleaned with a sterile saline solution. Initially, the inoculum starter were ready had around 4 x10<sup>7</sup> cfu/ml of cells. From Chr. Hansen, (Copenhagen, Denmark) probiotic strain of *L. plantarum* strain was obtained.

#### Preparation of probiotic pomegranate juice

Pomegranate juice extract 100 mL was pasteurized at 80 °C for 5 minutes, after cooling to 30 °C inoculated with 1 % *L. plantarum* under aseptic conditions and 24 hour incubation at 37 °C. Later kept at 4 °C in refrigerator until use after fermentation, no sugar or preservatives were used (Mamta et al., 2018).

#### Methods

1-Assessment of total phenolic concentration for pomegranate peel, pomegranate juice, and probiotic pomegranate juice extracts.

by Folin-Ciocalteu Colorimeter method, spectrophotometrically at 765nm. Gallic Acid Equivalents (mg GAE/g) were used to measure the total phenolic content (Mamta and Sharma, 2017).

2- Assessment of total Flavonoids content of pomegranate peel, pomegranate juice and probiotic pomegranate juice extracts. Total flavonoids content was measured spectrophotometrically at 510nm by the method of (Zhishen et al., 1999). Expressed as mg Quercetin Equivalent /g of dry weight sample on a calibration plot.

3- Assessment of antioxidant properties of pomegranate peel, pomegranate juice and probiotic pomegranate juice extracts. The 2, 2-diphenyl-1-picrylhydrazyl (DPPH) radical was used to make the determination Brand (Williams et al., 1995).

4- Phenolic and flavonoid fractionation and identification using high-performance liquid chromatography (HPLC) the polyphenolic compounds determined by (Pinto et al., 2008) but the flavonoids described by (Pirjo Mattila et al., 2000).

5- Determination of vitamins E and K detected by (Plozza et al., 2012). Ascorbic acid (Vitamin C) studied with (Romeu-Nadal et al., 2006).

6- Antimicrobial activity evaluation of pomegranate peel extracts, pomegranate juice and probiotic pomegranate juice. It was done using the well-agar diffusion method by (Perez et al., 1990).

The microorganisms used in this study included bacteria gram positive *Bacillus cereus* ATCC 33018 and *Staphylococcus aureus* ATCC 25923 and, bacteria gram negative *Pseudomonas aeruginosa* (ATCC 27853) and *Salmonella typhimurium* (ATCC 20231) and *Escherichia coli* (ATCC 25922). foodborne fungi such as *Aspergillus niger* (Rizk et al., 2009), *Fusarium verticillioides* (Sacc.) Nirenberg, and *Candida albicans* (CAIM-22) was acquired at Ain-Shams University in Cairo, Egypt, from MIRCEN (Microbiology Research Center). The bacterial and fungus cultures were diluted with sterile saline to nearly  $1.0 \times 10^5$  CFU/mL to make the spore suspension.

7- Antidiabetic *in Vitro* by inhibition of  $\alpha$ -Amylase Used the 3,5-dinitrosalicylic acid (DNSA) tech-

nique to conduct the experiment by (Wickramaratne et al., (2016).

8- Experimental Animals and diet

In these investigation 36 male albino rats of Wister strain weighing at  $150.00 \pm 20$  g was used. The rats were supplied from the National Research Center (NRC), Giza, Egypt. All experiment rats were done under the animal research ethics applied in the NRC up to 6 weeks (Abo-Taleb et al., 2017). Hypocholesterolemic ATOR (Atorvastatin) 20mg was obtained from the (EIPICO) Egyptian International Pharmaceutical Industries Company (10<sup>th</sup> of Ramadan city, Egypt) at a dose of 0.18 mg/Kg body weight /day (Guerin et al., 2000). Rats were given a standard diet and water (Reeves et al., 1993). Six groups of six rats each were created from the 36 rats. Group (1): Control rats (normal stander diet). Group (2): (positive control) High Fat Diet +1 % cholesterol.

Group (3): positive control treated with extract of pomegranate peel 300 mg /kg BW per day. Group (4): positive control treated with 1ml/day/rat pomegranate juice. Group (5): positive control treated with 1ml /day/rat from probiotic pomegranate juice. Group (6): positive control treated with ATOR standard drug. Daily checks on the rats' health revealed no detrimental developments during the period of the study. The dose for groups (3,4,5 and 6) take orally by stomach tube.

### Biochemical Examination

The blood samples from overnight-fasted rats were centrifuged for 10 minutes at 4 °C at 3000 rpm to get clear serum, which was then frozen at -18°C. Assay kits obtain from (Bio diagnostic, 29 Tahreer st., Dokki, Giza, Egypt).

Serum analysis: detection of serum cholesterol by (Richmond, 1973), triglycerides by (Fossati and Prencipe, 1982), and LDL and HDL by (Castelli and Levitar, 1977). (Reitman and Frankel's, 1957) method is used for liver enzymes tests of (AST) Aspartate amino transferase and (ALT) Alanine amino transferase, Alkaline phosphatase (ALP) by (Belfield and Goldberg, 1971).

### Histopathological examination

The experiment's effects on the liver and heart were assessed. At Cairo University's The Pathology Department at the University of Veterinary Medicine in Giza, Egypt prepared livers for histological analysis. Sections of liver samples that were placed in paraffin and were 3 to 5 mm thick were used. Hematoxylin-eosin (H&E, 400X) was stained the sections for light microscopy analysis

(Hirsch et al., 1997).

### Statistical analysis

Using Minitab Statistical Software version 15, one analysis of Variance (ANOVA) was performed on the data (Minitab Inc, State College, USA). The information presented in each table as mean  $\pm$  SD. (AboAllam, 2003).

**Table 1. Formulation composition of stander basal and High Fat diets (g/100g diet)**

Components	Stander basal diet (negative control )	Positive control (High fat diet + 1 % cholesterol)
Casein(90% protein)	18	18
Starch	62	47.62
Corn oil	10	8
Vitamin mixture(AIN-93-VX)	1	1
Mineral mixturex (AIN-93-MX)	4	4
Cellulous	5	5
Beef fat	_____	15
Cholesterol	_____	1
Choline bitartrate	_____	0.20
Cholic acid	_____	0.18

According to Terpstra et al., (2002)

### 3. Results and Discussion

Pomegranate peel, seeds, and arils are three different anatomical origins that can be recognized. Pomegranate juice, which can be made from the fruit's whole pomegranate or its arils (seed + juice). Results in Table 2. showed that about  $42.63 \pm 3.61\%$  of the fruit weight in regard to peel  $57.37 \pm 3.61\%$  arils,  $16.71 \pm 0.78\%$  seeds and  $40.66 \pm 3.58\%$

Juice. (Karimi et al., 2017) reported that when compared to the edible portion of the fruit, which was made up of 10% seeds and 40% arils, the peel, which form 50% of the fruit's non-edible portion, is distinguished by its antioxidant ability. The total phenolic concentration (mg Gallic acid/gm), total flavonoid contents (mg Quercetin /gm) and Antioxidants activity.

**Table 2. Percentage parts content of pomegranate fruit**

Parts of pomegranate fruit	Percentage content (%)
Peel	$42.63 \pm 3.61$
Arils (seed +juice)	$57.37 \pm 3.62$
Seeds	$16.71 \pm 0.78$
Juice	$40.66 \pm 3.58$



In Table 3. the total phenolic content of pomegranate peel was great than pomegranate juice and probiotic pomegranate juice, according to the data. The most significant results obtained in the extracts of pomegranate peel, probiotic juice and juice which contained  $665.64 \pm 1.02$ ,  $345.30 \pm 0.414$  and  $219.42 \pm 1.13$  mg GAE/g respectively. The pomegranate peel has a high phenolic content because it is full of ellagitannins, which are phenolic chemicals. Ellagitannins' polar nature prompted their extraction in ethanol, methanol, and water, two polar solvents (Hasan et al., 2018). As evidenced in Table 3. that the concentration of total flavonoid content the best amount of flavonoids were present in extracts of pomegranate peel, probiotic juice and juice. the results agree with (Kennas, 2019) which showed The greatest amounts of flavonoids  $53.85 \pm 1.95$ ,  $52.68 \pm 1.97$   $\mu\text{g QE/mg}$  respectively, in ethanolic and combination water/methanol (50:50) extracts, were little different from one another ( $P > 0.05$ ). Also detected that the total phenolic content in pomegranate peel extract ranged from  $242.05 \pm 7.99$  to  $638.17 \pm 10.59$  mg GAE/g but flavonoid content of pomegranate peel extracts varied  $63.85 \pm 1.9$  ( $\mu\text{g QE/mg}$  of dry extract). In comparison to other extracts, the pomegranate peel extracts demonstrated stronger antioxidant effects. As a result of fermentation, the radical scavenging rate increased (up to 40%), the highest antioxidant

rate of probiotic pomegranate juice more than and unfermented pomegranate juice (Pontonio et al., 2019). Because of the effect of sunlight and variances in local production areas on the formation of phenolic compounds and antioxidant properties. (Elfalleh et al., 2012). The results are in coincide with the (Alyaa et al., 2020) who reported that the order of Phenolic compound , flavonoid and antioxidants contents were as follows pomegranate peel extract > probiotic pomegranate juice > more than its juice. (Gözlekçi et al., 2011) reported that the greatest amount of total phenolic concentration were detected in extract of pomegranate peel more than its juice .

(Abdel-salam et al., 2018) studied that probiotic juice of pomegranate was great in total phenolic concentration, antioxidant rate more than control juice. Also in Table (3) Pomegranate peel had the highest vitamin C content ( $37.025 \pm 0.02 \text{mg}/100\text{g}$ ), while the pomegranate juice ( $11.044 \pm 0.001 \text{mg}/100\text{g}$ ) but the probiotic pomegranate juice  $14.021 \pm 0.005 \text{mg}/100\text{g}$ . (Vahid Akbarpour et al., 2009) found that the pomegranate juices vitamin C content also ranged between 9.68-17.45 mg/100 ml. the Vitamin A of Manfaloty peel powder is 0.593 mg/100g. Pomegranate juice had the highest vitamin K (18.568 mg/100g). (Pranav et al., 2017) who detected that peel of pomegranate contained ( $14.06 \pm 0.08 \mu\text{g/gm}$ ) vitamin A.

**Table 3. The total phenolic, total flavonoid, Antioxidants activity and Vitamines content for pomegranate peel, pomegranate juice and probiotic pomegranate juice.**

Samples	pomegranate peel	pomegranate juice	probiotic pomegranate juice
Total phenol (mgGE/g)	$665.64 \pm 1.02$	$219.42 \pm 0.133$	$345.30 \pm 0.414$
Total flavonoid (mg QE /g)	$66.20 \pm 0.451$	$31.2 \pm 0.442$	$48.2 \pm 0.432$
Antioxidant activity %	$93.82 \pm 0.202$	$51.16 \pm 0.161$	$60.44 \pm 0.102$
Vitamines			
Vit. C (L-scarobic acid) mg/100g	$37.025 \pm 0.062$	$11.044 \pm 0.001$	$14.021 \pm 0.005$
Vit. E ( $\alpha$ -Tochoferol) mg/100g	$0.012 \pm 0.001$	$0.017 \pm 0.007$	$0.076 \pm 0.00$
Vit.K (Phylloquinone) mg/100g	$3.032 \pm 0.002$	$18.568 \pm 0.007$	$20.068 \pm 0.004$

Data represent mean  $\pm$  SD, are not significant difference at  $P < 0.05$ .

Table 4. presents that composition of polyphenolic compounds of pomegranate peel powder, probiotic pomegranate juice and juice which were fractionat- ed into 19 different components, respectively by HPLC.

**Table 4. Composition of polyphenolic and flavonoid concentration of pomegranate peel powder, pomegranate juice and probiotic pomegranate juice by HPLC**

Phenolic compounds	Pomegranate peel ( $\mu\text{g/g}$ )	pomegranate juice ( $\mu\text{g/g}$ )	probiotic pomegranate juice ( $\mu\text{g/g}$ )
Gallic acid	22.2	3.5	4.8
Pyrogallol	1609.2	144.2	166.1
4-Amino-benzoic acid	18.4	2.2	4.2
Protocatechuic acid	282.3	76.2	91
Catechein	1766.1	530.1	607
Chlorogenic acid	413.3	43.5	56.4
Catechol	9.3	1.4	2.2
Caffeine	113.1	23.3	38.3
P-OH- benzoic acid	165.02	56.1	71
Caffeic acid	1.3	0.002	0.01
Vanillic acid	97	2.6	4.7
Ferulic acid	38	8.1	10.04
Iso-Ferulic acid	2.1	0.42	0.58
Ellagic acid	2039	43.3	60.2
Benzoic acid	223	20.1	33.6
Alpha-Coumaric acid	4.2	0.3	0.51
3,4,5-methoxy-cinnamic acid	30.1	3.6	4.9
Coumarin acid	24.6	nd	nd
Salicylic acid	28.5	0.008	0.02
	Flavonoid compounds mg/100g		
Apig.6-rhamnose8-glucose	20.412	2.063	3.145
Naringin	26.259	2.074	3.011
Rutin	37.654	2.659	4.234
Quercetin-3-o-glucoside	4.738	0.385	0.512
Apig.7-o-neohespiroside	69.995	3.362	5.427
Quercetin	60.674	1.456	2.321
Rhamentin	4.150	0.580	0.644
Apigenin	1.874	0.085	0.011
Kampferol	1.213	0.136	0.266
Naringenin	2.025	0.116	0.209

The percentage composition of polyphenolic compounds for pomegranate peel powder, that Ellagic acid had the highest composition for peel powder (2039  $\mu\text{g/g}$ ), while caffeic acid for Manfaluty peel powder showed the lowest value (1.3  $\mu\text{g/g}$ ). (Seba et

al., 2021) showed that ellagic acid content and catechin are the greatest amount in phenolic compounds, ellagic acid was major phenolic in Pomegranate peel (1034.7  $\mu\text{g/g}$ ), and Pomegranate juice (26.5  $\mu\text{g/g}$ ). Also in Table 4., quantitative

and qualitative analysis of flavonoids by HPLC were fractionated into 10 different components, Hesperitin, Luteo.7-glucose and Hesperidine were not found.

### Antimicrobial activity of pomegranate peel, probiotic pomegranate juice and juice

Pomegranate has an antimicrobial effects on different foodborne pathogens, which has an inhibit effect towards bacteria gram negative, gram-positive, fungi and yeast. The data of antimicrobial testing are showed in the Table 5. pomegranate showed the good results against different microbes ,the extracts of pomegranate peel powder has maximum inhibition zone than pomegranate juice, the phenolic compounds were more stable in solvent extracts than water extract. Similarly to (Parimala et al., 2018) shown that the extracts obtained from the solvent extracts ethanol, methanol, acetone higher than water extract. The extract of pomegranate peel produced greatest inhibition zones against *Staphylococcus aureus* 34 mm but the inhibition zone formed with, *Fusarium verticillioides* , *Escherichia coli* , *Candida albicans*, *Bacillus cereus* , *Salmonella typhimurium*, *Pseudomonas aeruginosa* and *Aspergillus niger* are 31, 30, 29, 28, 27, 25 and 24mm, respectively.

As shown in Table 5. the probiotic pomegranate juice was higher than juice in the diameter of inhibition zone. The result agrees with (Abdel-Aziz et al., 2021) studied that peel extracts of pomegranate exhibit antibacterial inhibition against the tested *E. coli* isolate, with the aqueous extract of pomegranate peel having a greater impact more than methanol extract on the tested *E. coli* strain.

(Dimitra et al., 2015) demonstrated that the ellagitannins and flavonoids found in pomegranate juices and fruit peels contributed to the substantial antibacterial activity of these matrices. Pomegranate juices showed considerable antibacterial inhibition and antifungal activity, which could increase their shelf time and stop deterioration brought on by the enzymatic activity of the microorganisms.

Stefanou et al., (2020) demonstrated that the juice, extracts, and isolated chemicals from pomegranates are effective against a variety of infections, with results that are frequently better than those of conventional antibiotics.

The data Similarly to Saad et al., (2010) and Priyanka Kesur et al., (2016 ) reported that comparing peel extract to other juice and seed extracts, peel extract has the perfect antibacterial activity. The strongest antibacterial activity toward *Staphylococcus aureus* was observed among the chosen bacterial and fungal cultures, and high antifungal activity toward *Aspergillus niger*.

(Akhtar et al., 2015) Based on their ability to precipitate cell membrane proteins with bacterial which causes lysis of bacterial, it has been hypothesised that polyphenolic compounds play an important part in the inhibition of microorganisms. These compounds may also stop microbial enzymes by acting with proteins by nonspecific interactions or by precipitating with sulfhydryl groups.

It has been demonstrated that pomegranate juice and peel have an unusually wide range of powerful antimicrobial and antiparasitic action (Celiksoy et al., 2021). The reduced pH and higher amount of organic acids in the fermented fruit juices were primarily responsible for the increased antimicrobial activity (Hao et al., 2020).

### In Vitro Antidiabetic Activity

By releasing oligosaccharides and monomeric molecules from dietary complex carbs, the alpha-amylase enzyme increases postprandial glucose levels in diabetics. Preventing the first enzyme needed for starch breakdown,  $\alpha$ -amylase and can slow down the rate at which blood glucose is released when a person consumes carbohydrates (Ponnusamy, et al., 2011).

In Table 6. Antidiabetic activity of pomegranate peel, juice and probiotic fermented juice was described by 3,5- dinitrosalicylic acid assay. From the results, the pomegranate peel have the highest of  $\alpha$ -amylase inhibition (97.8±0.002%) compared to all the others, which is associated with lower IC<sub>50</sub> value. (Seba et al., 2021) reported that the

*Punica granatum L.* peel has great carbohydrates inhibitory activity more than juice. Anthocyanin pigment, phenolic concentration found in the food chain from different fruits and vegetables and fermentation have been stopped and block digestive enzymes (Mojica et al., 2015).

Probiotic pomegranate juice is more effective and inhibition more than juice in comparison to the fresh juice, fermentation with various LAB strains of mango juice results in considerably higher inhibitory activities of  $\alpha$  amylase ( $p \leq .05$ ), (Florence et al., 2021).

**Table 5. shows the antimicrobial activity of extracts of pomegranate peel extract, pomegranate juice and probiotic pomegranate juice.**

Type of Extract	Antibacterial activity (diameter of inhibition zone mm)					Antifungal activity		
	Gram positive		Gram negative			fungi	yeast	
	<i>Staphylococcus aureus</i>	<i>Bacillus cereus</i>	<i>Pseudomonas aeruginosa</i>	<i>Salmonella typhimurium</i>	<i>Escherichia coli</i>	<i>Aspergillus niger</i>	<i>Fusarium verticillioides</i>	<i>Candida albicans</i>
Peel extract	34	28	25	27	30	24	31	29
Pomegranate juice	26	18	20	19	20	17	22	18
Probiotic pomegranate Juice	28	20	21	21	22	18	23	20

**Table 6. Inhibition of  $\alpha$ -amylase and IC<sub>50</sub> values in pomegranate peel, pomegranate juice and probiotic pomegranate juice.**

Samples (Conc. $\mu$ g/ml)	Amylase Inhibition %		
	Pomegranate peel	pomegranate juice	Probiotic pomegranate juice
1000	97.8 <sup>a</sup> ±0.002	64.3 <sup>c</sup> ±0.003	77.9 <sup>b</sup> ±0.009
800	90.4 <sup>a</sup> ±0.002	55.4 <sup>c</sup> ±0.002	68.3 <sup>b</sup> ±0.008
600	80.6 <sup>a</sup> ±0.0017	47.2 <sup>c</sup> ±0.003	59.9 <sup>b</sup> ±0.003
400	71.3 <sup>a</sup> ±0.006	39.6 <sup>c</sup> ±0.003	53.2 <sup>b</sup> ±0.004
200	58.5 <sup>a</sup> ±0.011	31.6 <sup>c</sup> ±0.004	48.8 <sup>b</sup> ±0.007
100	44.2 <sup>a</sup> ±0.011	22.0 <sup>c</sup> ±0.009	39.8 <sup>b</sup> ±0.003
80	35.4 <sup>a</sup> ±0.005	13.7 <sup>c</sup> ±0.005	31.7 <sup>b</sup> ±0.008
40	27.9 <sup>a</sup> ±0.009	5.4 <sup>c</sup> ±0.004	22.4 <sup>b</sup> ±0.017
20	18.7 <sup>a</sup> ±0.006	3.0 <sup>c</sup> ±0.002	12.5 <sup>b</sup> ±0.008
10	10.6 <sup>a</sup> ±0.002	1.1 <sup>c</sup> ±0.008	4.5 <sup>b</sup> ±0.001
Control 100%	0.0±0.037	0.0±0.037	0.0±0.037
IC <sub>50</sub>	34.43	84.8	40.04

Results represent mean  $\pm$  SD, the Identical letters in each Column are not significant at  $P < 0.05$ .

### Effect of pomegranate peel extract, pomegranate juice and probiotic juice on lipid profile and liver enzymes in hypercholesterolemic rats induced by HFD

Table 7. showed that the comparison effect be-

tween the pomegranate peel extract, pomegranate juice and probiotic pomegranate juice on high fat diet rats and negative group (basal diet), significant difference for all results. The group of rats takes 300mg of pomegranate peel extract was better than



other and its data near the reference drug (ATOR) which increases total cholesterol, LDL and triglyceride but raising the rate of HDL. The data agree with (Abo-Taleb, et al., 2017) which study the improvement action of juice and peel pomegranate extract on the total cholesterol, LDL, HDL and triglycerides in as hypercholesterolemic. Also (Lotfy and Alamri, 2019) reported that consumption of pomegranate peel which considered a high fiber source effect on anti-oxidative activities. It is used for weight loss and to improve the lipids profile. Also natural agent against obesity and high lipid levels. In Table 7. significant increase of ALT, AST and ALP values in diet high with cholesterol and fat in comparison to negative control rats while significant reduction was detected in the several parameters after 6 weeks of treatment with pome-

granate peel extract, probiotic juice, pomegranate juice and reference drug post induction with high fat diet in comparison to high fat diet (positive control group). These outcomes are consistent with earlier research that discovered rats fed pomegranate peel extract up to six weeks had significantly lower value of the ALT and AST enzymes. (Mansour et al., 2020) showed that significantly improves the liver markers after receiving gradient doses of. Pomegranate peel and juice also showed a substantial reduction in AST and ALT in rats. (Alireza et al., 2014) obtained that rats fed a high-lipid diet, peel extract of the pomegranate dramatically lower serum levels of lipid composition profile and hepatic enzyme, while raising serum HDL-C in comparison to stander control.

**Table 7. Activity of peel extract, juice and probiotic juice of pomegranate on lipid composition profile and liver enzymes of hypercholesterolemic rats.**

Parameter	Negative control	Positive (HFD) control	(HFD) + 300 mg Peel extract	(HFD) +1ml juice	(HFD) + 1ml Probiotic juice	HFD + ATOR
Total cholesterol (mg/dl)	92.10 <sup>e</sup> ± 0.33	157.20 <sup>a</sup> ± 1.04	95.70 <sup>d</sup> ± 0.32	100.80 <sup>b</sup> ± 0.45	97.83 <sup>c</sup> ± 0.05	96.07 <sup>d</sup> ± 1.08
LDL(mg/dl)	29.83 <sup>f</sup> ± 0.65	107.10 <sup>a</sup> ± 0.09	36.07 <sup>d</sup> ± 0.55	50.07 <sup>b</sup> ± 0.26	41.03 <sup>c</sup> ± 0.35	34.60 <sup>e</sup> ±
HDL(mg/dl)	53.33 <sup>a</sup> ± 0.23	31.05 <sup>f</sup> ± 1.00	50.71 <sup>c</sup> ± 0.65	43.77 <sup>c</sup> ± 0.02	48.74 <sup>d</sup> ± 0.22	51.5 <sup>b</sup> ± 0.51
Triglycerides (mg/dl)	46.06 <sup>f</sup> ± 0.24	93.11 <sup>a</sup> ± 0.76	48.06 <sup>d</sup> ± 0.50	55.07 <sup>b</sup> ± 0.97	49.05 <sup>c</sup> ± 0.51	47.05 <sup>c</sup> ± 0.01
liver function enzymes						
ALT (U/l)	50.06 <sup>f</sup> ± 0.82	90.23 <sup>a</sup> ± 0.34	51.21 <sup>c</sup> ± 0.65	59.06 <sup>b</sup> ± 0.82	55.06 <sup>c</sup> ± 0.13	52.36 <sup>d</sup> ± 0.12
AST (U/l)	43.30 <sup>f</sup> ± 0.57	70.01 <sup>a</sup> ± 0.20	55.65 <sup>c</sup> ± 0.22	60.65 <sup>b</sup> ± 0.22	57.65 <sup>c</sup> ± 0.22	56.65 <sup>d</sup> ± 0.65
ALP (U/l)	70.90 <sup>f</sup> ± 4.00	136.84 <sup>a</sup> ± 1.10	85.08 <sup>e</sup> ± 0.42	95.33 <sup>b</sup> ± 1.06	90.20 <sup>e</sup> ± 1.29	88.14 <sup>d</sup> ± 0.15

Data represent mean ± SD, the identify letters in each column are not significant at P < 0.05.

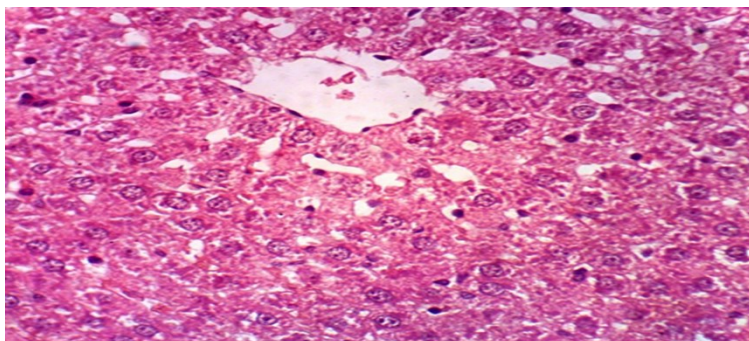
### Histological Assessment

In Figure 1. no histopathological modification in the negative control rats due to the parenchyma's major vein and bordering hepatocytes had their usual histological structure, whereas the blood vessels of revealed . But in positive control high-fat diet-fed rats showed in Figure 2. addition to substantial dilatation and congestion in the portal vein, the histology of blood vessels in showed many intracyto-

plasmic micro fat vacuoles in the hepatocytes as a fatty change. After being treated with pomegranate peel extract, high-fat diet rats revealed normal liver lobule histology with low inflammatory cell infiltration in the blood vessels in Figure 3. In both Figures 4 and 5 treated of the portal region which had fibrosis and dilatation in the portal vein by pomegranate juice 1ml/day/rat and 1ml /day/rat from probiotic pomegranate juice .

In the Figure 6. showed normal histological structure of the hepatic lobule and normal histological structure of the central vein when treated with the reference medication ATOR in high lipid diet. The results are in agreement with (Alireza et al., 2014) which reported that the pomegranate peel extract

formed treatment and reduced hepatic damage in rats fed a high-lipid diet in comparison to a negative control. Our results are in agreement also with the results of (Abo-Taleb et al., 2017) and (Binmowyna et al., 2021).



**Figure 6. showed normal histological structure when treated with the reference medication ATOR in high lipid diet.**

#### 4. Conclusion

Pomegranate bio waste, pomegranate juice and probiotic juice are a good source of bioactive compound and antioxidant activity, also have strong antimicrobial activity. The benefits of drinking non-dairy fermented beverages for health are confirmed by probiotic juice, which vegetarians and those with high cholesterol can also use., the Pomegranate is a potential treatment alternative as an anti-hyperlipidemic drug, but more clinical research is required to validate this.

#### References

Abdel-Aziz M.A.E., Hazaa M.M., Hadeer Y.A. and Mervat G.H. (2021). Antibacterial Potential of Pomegranate Peel Extracts on *Escherichia coli* Isolated From Benha Hospital in Egypt. *Benha Journal of Applied Sciences (BJAS)* .6:(3) Part (1), 61-64. <http://bjas.journals.ekb.eg>.

Abdel-salam, F.F., Moharram Y.G. and El-Zalaki, E.M. (2018). Wonderful Pomegranate (*Punica granatum L.*) Juice Wastes Extract as a Food preservative .*Egypt. J. Food Sci.* 46, 101- 111.

Abo-Allam R. M. (2003). Data statistical analysis using the SPSS program.1st, ed., a publication for Universities, Cairo.

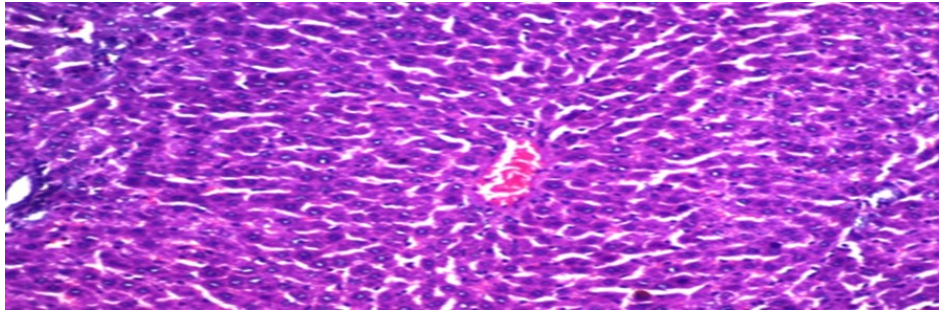
Abo-Taleb H.M., Mona M. A.M. , Nadia A. R. S. and Mahmoud A.M.S. (2017). Evaluation of pomegranate (*punicagranatum L.*) Freeze dried juice and peel powder extracts as antidiabetic and antihyperlipidemic agents in rats. *IOSR Journal of Pharmacy and Biological Sciences (IOSR-JPBS)* e-ISSN: 2278-3008, p-ISSN:2319-7676. 11, ( 6 )Ver. II, 53-64.

Akhtar S., Ismail T., Fraternal D. and Sestili P. (2015). Pomegranate peel and peel extracts: Chemistry and food features. *Food Chemistry*, 174,417- 425.

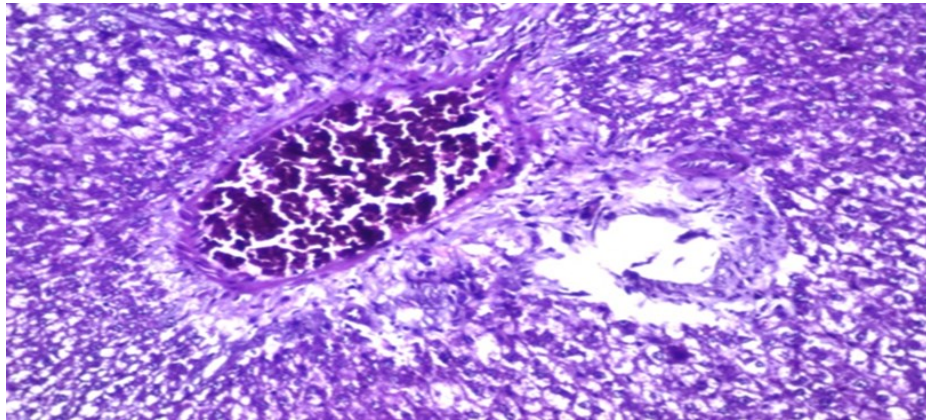
Alkhatib M.,Fayad, C., BadranA., Hamade K.,Daou A., Baydoun, E.and Hijazi A.(2022). Preventive and Therapeutic Effects of *Punica granatum* (Pomegranate) in Respiratory and Digestive Diseases: A Review. *Appl. Sci.*, 12, 12326. <https://doi.org/10.3390/app122312326>.

Alireza S., Maryam E., Ali I. Kavgani R. Ghahramani S. and Ali A.(2014). Lipid Lowering Effect of *Punica granatum L.* Peel in High Lipid Diet Fed Male Rats. *Evidence-Based Complementary and Alternative Medicine*. 2014: 432650,1-5. <http://dx.doi.org/10.1155/2014/432650>.

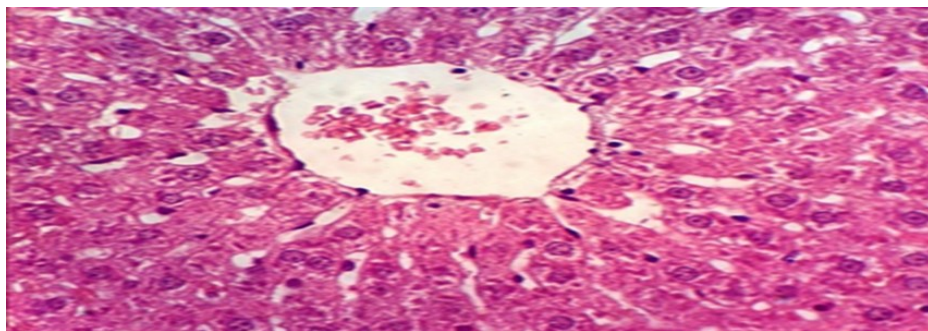




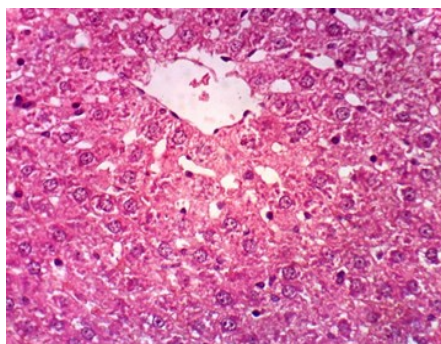
**Figure 1.** The negative control rats revealed no histopathological modification (H & E X 400) .



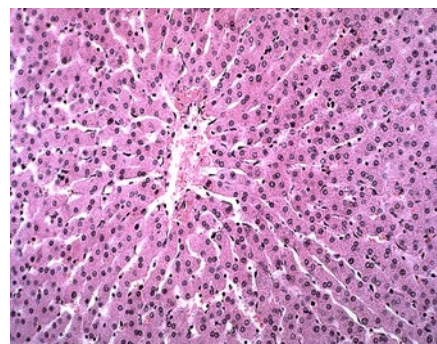
**Figure 2.** Positive control (high-fat diet-fed rats) hepatic lobule (H & E X 400)



**Figure 3.** After being treated with pomegranate peel extract 300 mg Peel extract



**Figure 4.** Liver of rats from positive control+1ml /day /rat pomegranate juice



**Figure 5.** Liver of rats from positive control+1ml /day /rat probiotic pomegranate juice.

- Alyaa M. E., Nihal S. and Hanan A. S.(2020). Chemical Studies, Antioxidant and Antimicrobial Activities of Pomegranate and it's possible use some food production. Master Thesis, Organic Chemistry, Faculty of Science, AL-Azhar University .
- Amri M. R. K. , Mohamed S. , Wafa K. , Mouna T. , Fatma A. and Mohamed H. (2020) . Anti-diabetic effects of pomegranate extracts in long-term high fructose-fat fed rats. *Clinical Phyto-science* 6:55. <https://doi.org/10.1186/s40816-020-00202-y>.
- Andi Alfira R. F., Muntholib D. and Subandi (2020). In vitro and In silico Analysis of Pomegranate (*Punica granatum L.*) Fruit Powder as Pancreatic Lipase and  $\alpha$ Amylase Inhibitor. *Journal of Physics. Conf. Ser.* 1665 (1):012004,1-8.
- Belfield A. and Goldberg D. M. (1971). "Colorimetric Determination of Alkaline Phosphatase Activity". *Enzyme*, 12 (5): 561-573.
- Binmowyna M.N., Nora A. , Ahmad T.A., Muneer M. A., Ekram A. A.(2021). Hypolipidemic and antioxidant effects of the juice and water seed extracts of two pomegranate species in high-cholesterol diet fed rats. *Food Sci. Technol, Campinas*, 41(2): 732-740.
- Brand-Williams W., Cuvelier M. and Berset C. (1995). Use of a free radical method to evaluate antioxidant capacity. *Food Science and Technology*, 28(1): 25–30.
- Castelli and Levitra (1977). Atherogenic indices, *J. Curr Presc .*, 39.
- Celiksoy V., Rachael L., Alastair J. S., Ryan M. and Charles M. H. (2021). "Synergistic In Vitro Antimicrobial Activity of Pomegranate Rind Extract and Zinc (II) against *Micrococcus luteus* under Planktonic and Biofilm Conditions" *Pharmaceutics* ,13(6):851. <https://doi.org/10.3390/pharmaceutics13060851>.
- Devatkal S. K. and Naveena B. M. (2010). Effect of salt, kinnow and pomegranate fruit by-product powders on color and oxidative stability of raw ground goatmeat during refrigerated storage. *Meat Science*, 85(2): 306–311.
- Dimitra Z. L., Vassilia J. S., Panagiotis Z. and Charalampos P.(2015). Comparison of the Antioxidant and Antiradical Activity of Pomegranate (*Punica Granatum L.*) by Ultrasound-Assisted and Classical Extraction .*Analytical Letters*, 37-41. DOI: 10.4172/2329-6836.1000332.
- Elzoghbiy A. S., Mahmoud M. M.Mohammed H. Elhaw and Alsayed E. M. (2022). Phytochemical Hytochemical Analysis OF Pomegranate Peel Extract (PPE) With The Evaluation of Its Efficiency AS Anti-rancidity OF Oils, Anti-Bacterial And Anti-Candidn Agent. *Al azaher Medical Journal* , 51: (3) . 1851-1866.
- Elfalleh W., Nasri N., Marzougui N., Thabti I., M'Rabet A., Yahya Y., Lachiheb B., Guasmi F. and Ferchichi A. (2012). Total phenolic contents and antioxidant activities of pomegranate peel, seed, leaf and flower. *Journal of Medicinal Plants Research* . 6(20 .4730-4724 ,(DOI: 10.5897/JMPR11.995.
- Florence M. Mashitoa, Vimbainashe E. Manhivi, Stephen A. AkinolaC. G.,Fabienne R., Tinotenda S. and Dharini S. (2021). Changes in phenolics and antioxidant capacity during fermentation and simulated in vitro digestion of mango puree fermented with different lactic acid bacteria *J Food Process Preservation*,45 (11) , 15937.
- Fossati P. and Prencipe L. (1982). Serum triglycerides are determined calorimetrically with an enzyme that produces hydrogen peroxide. *Clin. Chem.* 28 (1): 2077 –2080.
- Gözlekçi S., Saraçoğ'Ö., Onursal E.and Özgen M. (2011). Total phenolic distribution of juice, peel, and seed extracts of four pomegranate cultivars. *Pharmacognosy Magazine*, 7, 161–164.
- Guerin M., Lassel T., Le Goff W., Farnier M. and John C. M. (2000). Action of Atorvastatin in Combined Hyperlipidemia. Preferential Reduction of Cholesteryl Ester Transfer From HDL to VLDL1 Particles. *J. Arteriosclerosis, Thrombosis and Vascular Biol.*, 20:189-198.



- Hao Z., Abdullah L. D., Minjie Z., Jun T., Tao L., Hui Z. and Fengqin F. (2020). Probiotic-fermented blueberry juice prevents obesity and hyperglycemia in high fat diet-fed mice in association with modulating the gut microbiota. *Food & Function*. 11, 9192–9207.
- Hasan A.M., Ali R. and Qaher M. (2018). Phytochemical Investigations of Pomegranate (*Punica granatum*) Rind and Aril Extracts and their Antioxidant, Antidiabetic and Antibacterial Activity *Nat Prod Chem Res*, 6(4):1-10.
- Hirsch C., Zouain C.S., Alves J. B. and Goes A.M. (1997). Induction of protective immunity and modulation of granulomatous hypersensitivity in mice using PIII, an anionic fraction of *Schistosoma mansoni* adult worm. *Parasitology*, 115: 21- 28.
- Hua Y., Guolin C., Jian L. and Encarna G.P. (2021). The production and application of enzymes related to the quality of fruit wine, *Critical Reviews in Food Science and Nutrition*, 61:10, 1605-1615, DOI: [10.1080/10408398.2020.1763251](https://doi.org/10.1080/10408398.2020.1763251)
- Karimi M., Rohollah S. and Jozef K.(2017). Review Pomegranate as a promising opportunity in medicine and nanotechnology .*Trends in Food Science & Technology*. 69( A) :59-73.
- Kennas A. and Amellal–Chibane H. (2019). Comparison of five solvents in the extraction of phenolic anti-oxidants from pomegranate (*Punica granatum L.*) peel. *The North African Journal of Food and Nutrition Research*: 3(5): 140-147 [https://doi.org/ 10.5281/zenodo.2567587](https://doi.org/10.5281/zenodo.2567587).
- Ko K., Younas D. and Alireza A. (2021). "Nutritional and Bioactive Components of Pomegranate Waste Used in Food and Cosmetic Applications: A Review" *Foods* 10(3): 657. [https://doi.org/ 10.3390/ foods10030657](https://doi.org/10.3390/foods10030657)
- Lotfy L.M. and Alamri E.S.(2019). The Impact of Pomegranate Peel-fortified Cupcakes on Weight Loss. *International Journal of Pharmaceutical Research & Allied Sciences*, 2, 8 (3):119-125.
- Mamta Thakur and Sharma R. K. (2017). Development of Probiotic Pomegranate Beverage and Its Physico-Chemical and Microbial Characterization. ISSN: 2320 – 7051 *Int. J. Pure App. Biosci.* 5 (1): 35-41.
- Mamta Thakur H. W., Deshpande M. and Bhate A. (2018). Investigation of Microbial, Physico-chemical and Color Properties of Probiotic Pomegranate Beverage during Storage. ISSN: 2319-7706 *Special Issue-:( 7) 638-650*.
- Mansour A. T., Mohamed M. M., Hossam S. E., and Ahmed E. S. (2020). "Synergism of Dietary Co-Supplementation with Lutein and Bile Salts Improved the Growth Performance, Carotenoid Content, Antioxidant Capacity, Lipid Metabolism, and Lipase Activity of the Marbled Spine foot Rabbit fish, *Siganus rivulatus*" *Animals* 10 (9): 1643. <https://doi.org/10.3390/ani10091643>.
- Min M., Bunt, C.R., Mason S.L.and Hussain M.A. (2019). Non-dairy probiotic food products: An emerging group of functional foods. *Crit Rev Food Sci Nutr.*; 59(16):2626-2641. doi: 10.1080/10408398.2018.1462760.
- Mojica L., Meyer A., Berhow M.A. and Mejía E.G.D.(2015). Bean cultivars (*Phaseolus vulgaris L.*) have similar high antioxidant capacity, in vitro inhibition of  $\alpha$ -amylase and  $\alpha$ -glucosidase while diverse phenolic composition and concentration. *Food Res Int* 69:38–48.
- Parimala C. and Jenifer M. (2018). Antibacterial activity of *Punica granatum* peel extract against selected ATCC pathogens *International Journal of Applied Research*,4(9): 74-77.
- Perez M. P. and Bazerque P.(1990). "An antibiotic assay by the agar well diffusion method", *Acta Biol. Med. Exp.*, 15, 113–115.
- Pinto M. S., Lajalo F. M. and Genovese M. I. (2008). Bioactive compounds and quantification of total ellagic acid in strawberries (*Fragaria x ananassa Duch*). *Food Chemistry*, 107: 1629-1635.
- Pirjo-Mattila J.A. and Jourma K.(2000). Determination of flavonoids in plant material by HPLC with diode-array and electro array detections. *J. Agric. Food chem.*48, 5834-5841.
- Plozza T., Trenerry V.C., and Caridi D. (2012). The simultaneous determination of vitamins A,



- E and  $\beta$ -carotene in bovine milk by high performance liquid chromatography-ion trap mass spectrometry (HPLC- M S n). *Journal of food Chemistry*, 134:559-563.
- Ponnusamy S., Ravindran R., Zinjarde S., Bhargava S. and Ravi Kumar A. (2011). Evaluation of traditional Indian antidiabetic medicinal plants for human pancreatic amylase inhibitory effect *in vitro*. *Evid Based Complement Alternat Med*. 2011,1-10. doi:10.1155/2011/515647.
- Pontonio E., Montemurro M., Pinto D., Marzani B., Trani A., Ferrara G., Mazzeo A., Gobetti M. and Rizzello C.G. (2019). Lactic Acid Fermentation of Pomegranate Juice as a Tool to Improve Antioxidant Activity. *Front. Microbiol.* 10:1550. doi: 10.3389/fmicb.2019.01550.
- Pranav D., Pathak S. A., Mandavgane B. and Kulkarni D. (2017). Fruit peel waste: characterization and its potential uses. *CURRENT SCIENCE*. 113,1-11.
- Priyanka K., Mayur G., Poonam B.C and Hirenghai V. (2016). Evaluation of Antimicrobial Properties of Peels and Juice Extract of Punica Granatum (POMEGRANATE). *IJRSI*, 3(5): 1-11.
- Reeves P.G., Nielsen F.H. and Fahey G.C. (1993). AIN-93 Purified Diets for Laboratory Rodents: Final Report of the American Institute of Nutrition Ad Hoc Writing Committee on the Reformulation of the AIN-76A Rodent Diet. *The Journal of Nutrition* 123(11):1939 –1951. <https://doi.org/10.1093/jn/123.11.1939>.
- Reitman S. and Frankel S. (1957). A colorimetric method for determination of serum glutamate oxaloacetate and glutamic pyruvate transaminase. *American Journal of Clinical Pathology*, 28: 56-58.
- Richmond N. (1973). Colorimetric method of determination of total cholesterol and high density lipoprotein cholesterol (HDLc). *Clin.Chem.*, 19: 1350-1356.
- Rizk, M. A, Khalil M.S. , Abu- Zaid A. and Magdi M. (2009). Evaluation of antimicrobial activity of stored fermented soybean products. *Bull.Fac.Sci., Cairo Univ.*, 77: 1-17.
- Romeu-Nadal M., Morera-Pons S., and Lopez-Sbater M.C.(2006). Rapid high-performance liquid chromatographic method for Vitamin C determination in human milk versus an enzymatic method. *Journal of chromatography B*, 830:41-46.
- Saad S. D., Mir N A., Hajera T. and Mazharuddin K. (2010). Studies on Antibacterial and Antifungal Activity of Pomegranate (*Punica granatum L.*) *American-Eurasian J. Agric. & Environ. Sci.*, 9 (3): 273-28.
- Seba A., Bülent B. and Mehmet K. (2021). Multivariate Analyses of the Antioxidant, Antidiabetic, Antimicrobial Activity of Pomegranate Tissues with Respect to Pomegranate Juice. *Waste and Biomass Valorization*. 12:5909–5921 <https://doi.org/10.1007/s12649-021-01427-9>.
- Stavros K., Ioanna M. and Stavros P. (2020). Assessment of Pomegranate Juice as an Alternative “Substrate” for Probiotic Delivery. *Recent Advances and Prospects. Fermentation*, 6, 24.
- Stefanou V., Tsakni A., Timbis D., Vougiouka P.A., Doumi I. and Maronikolaki (2020). Pomegranate as an Antibacterial Agent against Pathogens and at the same Time Advantageous to Beneficial Bacteria: A Review. *Int J Adv Res MicroBiol Immunol* 2(2): 1-13.
- Terpstra A.H., Lapre J.A., de Vries H. and Beynen A.C. (2002). The hypocholesterolaemic effect of lemon peels, lemon pectin, and the waste stream material of lemon peels in hybrid F1B hamsters. *Eur. J. Nutr.*, 41: 19-26.
- Vahid-Akbarpour; Khodayar-Hemmati and Mehdi-Sharifani (2009). Physical and Chemical Properties of Pomegranate (*Punica granatum L.*) Fruit in Maturation Stage *American-Eurasian J. Agric. & Environ. Sci.*, 6 (4): 411-416.
- Wickramaratne Nirmali, J. C. Punchihewa and D. B. M. Wickramaratne (2016). In-vitro alpha amylase inhibitory activity of the leaf extracts of *Adenantha pavonina*. *BMC Complementary and Alternative Medicine* 16:466; DOI 10.1186/s12906-016-1452-y.

Xiang Luo H. L., Zhikun W., Wen Y., Peng Z., Da C., Haiyan Y., , Krishna P., Diguang Z., Fuhong Z., Xiaocong X., Lina C., Qi W., Dan J. and Shangyin C.(2020). The pomegranate (*Punica granatum*L.) draft genomedissects genetic divergence between soft- and shard-seeded cultivars. *Plant Biotechnology* .18, 955–968.doi: 10.1111/pbi.13260.

Zhishen J., Mengcheng T. and Jianm (1999). The determination of flavonoid contents in mulberry and their scavenging effects on superoxide radicals. *Food Chemistry*, 64:555-559.